









Welcome

Graduate Research and Training Contacts Staff Student Liaison Committee/Officer

Welcome

We are delighted to welcome you to your postgraduate training programme at the Institute of Genetics and Cancer. On the following pages you will find information relating to the different programmes, timetable for the first 6 months, and the assessment timetable for the next 3 or 4 years.

As you probably know, we have a mixture of students on campus, some of whom are following a four year PhD programme with rotations and others who are starting a three year PhD in a specific lab, whilst others are studying for an MSc or MD. There are some teaching elements of the four year taught course that might be of interest to other students, for example covering different technologies, computer programming, aspects of clinical reserach and research ethics - these are shown in a detailed timetable. This teaching is only compulsory for the 4-year HGU students, but other students (and postdocs) are welcome to sign up and attend any sessions that you find useful, you might want to discuss your choice with your supervisor(s). We hope that the Graduate Research and Training environment will provide a useful framework for your studies. Please feel free to air your views, and to approach us about any issues you have, and help us to make the Institute a huge success!

Graduate Research and Training contacts

The Institute is made up of three centres, the MRC Human Genetics Unit (HGU), the Centre for Genomics and Experimental Medicine (CGEM) and the Cancer Research UK Edinburgh Centre (ECRC), each with their own Graduate Convenor. The Institute falls within the School of Molecular, Genetic and Population Health Sciences (you will need to know this School affiliation when you apply for Transkills courses amongst other things), and the SMGPHS is within the College of Medicine and Veterinary Medicine or CMVM. In the first instance you will mainly deal with your supervisors, Graduate Convenor or Nick Gilbert (Director of Graduate Research and Training for the Institute of Genetics and Cancer). You will also have a thesis committee (normally setup abour 10 weeks into your PhD) which will be made up of your supervisors, an external advisor and a committee Chair. Formal issues (interruption of studies and so on) are dealt with by the Director of Graduate Research and Training and the College PG Office.

Director of Graduate Research and Training, Institute of Genetics and Cancer: Professor Nick Gilbert

Graduate Convenor, CRUK Edinburgh Centre: Dr Susan Farrington

Graduate Convenor, MRC Human Genetics Unit: **Professor Ian Adams**

Graduate Convenor, Centre for Genomic & Experimental Medicine: Dr Kathy Evans

Director of PG Studies, College of MVM: Professor Paddy Hadoke

Staff Student Liaison Officer: Dr Catherine Naughton Dr Dasa Longman

Graduate Research and Training Administrator: Pauline McDonald

Graduate Research and Training Assistant: Alana Johnson

Staff Student Liaison Committee

At the Institute of Genetics and Cancer we are committed to ensuring a high-guality student experience. To ensure we are able to deliver this, and to "maximise our students' potential", we encourage students to communicate their views and suggestions to help influence any required changes to policies and procedures. The Institute Staff Student Liaison Committee (SSLC) meets biannually to discuss matters of mutual concern of staff and students. The SSLC is composed of student and staff representatives, and we strongly encourage students at any stage of their graduate degree to consider joining the SSLC. The current Staff Student Liaison Officers are Post-Doc Ed Jarman and (SSLO) Dr. Dasa Longman.



Ed Jarman

Ed is a postdoctoral research fellow with 6 years' experience working in the Liver Cancer and Regeneration Lab under Dr. Luke Boulter at the Institute of Genetics and Cancer. Ed previously completed his PhD at the IGC, and so brings experience of both the staff and student experience at the institute.



Dasa Longman

Dasa is a Senior Scientist in the lab of Professor Javier Caceres, MRC HGU, and has many years experience of formal and informal mentoring of PhD and undergraduate students.

Dasa and Ed together oversee the POGs induction events held during induction week for new PhD students, coordinate the 1st-year student journal clubs and organise the biannual SSLC meetings.

What to do if things go wrong

If you have a problem with your project and/ or supervisor, you should first try to resolve it between yourselves - it is important to keep lines of communication open where possible and not let things degenerate. If there is still a problem, then please seek advice - you should feel free to speak to your second supervisor, your thesis committee Chair, the Directors of the Graduate School or the PG Convenor for your building.

These conversations will be in confidence and a strategy will be devised to try and address any problems. Additional meetings of thesis committees can be arranged (subject to members' availability) if the student and/ or supervisors feel that this would help. If you are not happy with the outcome of frontline resolution (and on the rare occasions where a local resolution is not an appropriate early step) the University has procedures in place for dealing with complaints and the Institute of Genetics and Cancer adheres to these procedures rigorously. Details of these can be accessed through the CMVM Postgraduate Wiki which is also accessible from the Institute of Genetics and Cancer Graduate Research and Training web pages.

Meet the Team: PG Directors



Professor Nick Gilbert - Director of Graduate Research & Training MRC HGU/ Institute of Genetics and Cancer Email <u>Nick.Gilbert@ed.ac.uk</u> Telephone 0131 651 8551 Location: C3.21 Research Group www.ed.ac.uk/mrc-human-genetics-unit/research/gilbert-group



Dr Ian Adams - Graduate Convenor, MRC Human Genetics Unit Email <u>Ian.Adams@ed.ac.uk</u> Telephone 0131 467 8456 Research Group https://www.ed.ac.uk/mrc-human-genetics-unit/research/adams-group



Dr Susan M Farrington - Graduate Convenor, CRUK Edinburgh Centre Email <u>Susan.Farrington@ed.ac.uk</u> Telephone 0131 651 8632 Research Group www.ed.ac.uk/cancer-centre/research/farrington-group



Dr Kathy Evans - Graduate Convenor, CGEM Email <u>Kathy.Evans@ed.ac.uk</u> Telephone 0131 651 8747 Location: N2.09 Research Group www.ed.ac.uk/centre-genomic-medicine/research-groups/evans-group

Students and staff should contact their local Centre PG Director for academic support.

Administration Team



Pauline McDonald Alana Johnson

Email <u>student-admin@igc.ed.ac.uk</u> Telephone 0131 651 5771 Location: CG.11

Pauline and Alana manage the day-to-day administration of the Graduate Research and Training programme, and are based on the ground floor of the MRC Human Genetics Unit.

For queries related to Postgraduate Research and Training, Pauline and Alana provide support to prospective, on-programme and visiting students, as well as supervisors and academic staff. When appropriate, they will signpost students and staff to key central university services. Pauline and Alana work closely with Centre PG Directors to enhance the Student Experience and oversee the following areas of work:

- · Student Recruitment & Admissions
- Tier 4 Engagement & Monitoring process for international students
- · Visiting student admissions
- Manage Graduate Research and Training website in liaison with PG Directors
- · Coordinate teaching programme
- Organise student events e.g. Science at the Interface to Industry, Christmas lectures, John Inglis talks etc.
- Organise and minute Staff Student Liaison Committee (SSLC) / Postgraduate Studies Committee (PGSC)
- · Manage Student Social Media Platforms

Induction Week Teaching Timetable

Induction Week

Monday 9th Septe	mber				
09:30 - 10:00	PGR Director's welcome – Nick, Ian, Susan (Lecture Theatre)				
10:00 - 11:00	Health and Safety Induction - Iain Kennedy (Lecture Theatre)				
Break for coffee					
11:00 - 11:30	General Admin to Students Q&A - Pauline, Alana (Lecture Theatre)				
Break for lunch					
14:00 - 15:00	Good Practice in PhD Research - Grace, Maarten, Annabel (Lecture Theatre)				
15:30 - 17:00	POGS social - POGS Team (soft seating area of NUCLEUS)				
Tuesday 10th Sep	tember				
09:00 - 10:15	UoE ISG Digital Skills Team – Kenny, Riccardo (MEC Computing Lab 1)				
10:30 - 10:45	Edinburgh Innovations - Farai Munjoma (Lecture Theatre)				
10:45 - 11:15	HGU Students only - Intro to 4 year programme - Ian, Pauline (Lecture Theatre)				
Break for coffee					
11:30 - 12:00	CMVM Welfare Advice (Lecture Theatre)				
12:00 - 13:00	EUSA Advice Place with Pop-up - Clair Halliday (Lecture Theatre)				
Matriculation Old	College - Afternoon				
Matriculation Old Wednesday 11th S					
Wednesday 11th S	September				
Wednesday 11th S 09:00 - 09:30	September Library Services - Marshall Dozier (Lecture Theatre)				
Wednesday 11th S 09:00 - 09:30 09:30 - 10:00	September Library Services - Marshall Dozier (Lecture Theatre) Institute of Academic Services - Louise McKay (Lecture Theatre)				
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Monday 16th Sept	ember				
09:00 - 16:30	Project Organization management & data wrangling for genomics 1 - Graeme Grimes (MEC Computing lab 1)				
Tuesday 17th Sept	Tuesday 17th September				
09:00 - 16:30	Data Wrangling for Genomics 2 - Graeme Grimes (MEC Computing lab 1)				
Wednesday 18th S	September				
09:00 - 16:00	Introduction to Eddie - John ireland (MEC Computing lab 1)				
Thursday 19th Sep	ptember				
09:00 - 12:30	Intro to R and RStudio for Genomics 1 - Jing Su (MEC Computing lab 1)				
Break for lunch					
15:30 - 17:00	HGU rotation project talks 4 (E4.07)				
17:00 - 19:00	POGS Quiz (Nucleus)				
Friday 20th Septer	mber				
09:00 - 12:00	Intro to R and RStudio for Genomics 1 - Jing Su (MEC Computing lab 1)				
Break for lunch					
14:00 - 14:30	Student Health and Wellbeing - Andy Shanks (E4.07)				
14:30 - 15:00	Public Engagement and Communication - Faye Watson (E4.07)				
15:30 - 17:00	HGU rotation project talks 5 (E4.07)				
Monday 23rd Sept	ember				
09:00 - 16:30	Statistical thinking for public health, Simple Linear Regression - Hannes Becher (MEC Computing lab 1)				
Tuesday 24th Sept	tember				
09:00 - 16:30	Simple Linear Regression, Multiple linear regression for public health - Hannes Becher (MEC Computing lab 1)				
Wednesday 25th S	September				
09:00 - 16:30	Intro to Python - Murray Wham (MEC Computing lab 1)				
Thursday 26th Se	ptember				
09:00 - 16:30	Intro Git and GitLab - Graeme Grimes (MEC Computing lab 1)				
Friday 27th Septer	mber				
09:00 - 16:30	Intro to Genome Browsers and IGV - EBI Team / Gogo (MEC Computing lab 1)				

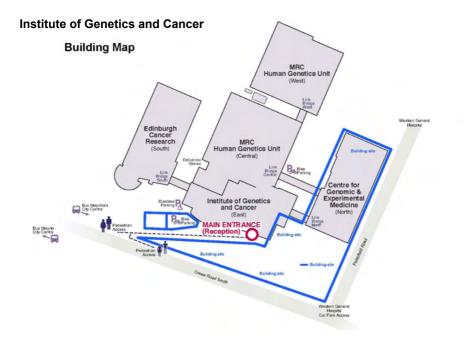
Monday 30th September HGU Students Start Rotation Project 1					
09:00 - 12:00	Reading and evaluating scientific literature - Ian Adams / Carolina Uggenti (E4.07)				
14:00 -15:30	How to duplicate your genome - Tom Deegan (E4.07)				
Monday 7th Octob	ber de la constant de				
09:30 - 11:00	Good Research Practice - Helen Nickerson (E4.07)				
14:00 - 15:30	Establishing mechanisms underlying genetic associations with complex traits and disease - Veronique Vitart and Chloe Stanton (E4.07)				
Thursday 10th Oc	tober				
16:00 - 18:00	POGS Career Event (E4.07)				
Monday 14th Octo	ber				
09:00 - 12:00	Experimental Design - Kevin Myant, Luke Boulter (E4.07)				
14:00 - 15:30	Critical Evalution Skills 1 (E4.07)				
Monday 21st Octo	ber				
09:00 - 12:00	Scientific Blogging - Kendal Flack - Online Workshop				
14:00 -15:30	Preventing Inherited Aneuploidies in the Mammalian Germline - Ian Adams (E4.07)				
Monday 28th Octo	ber				
09:00 - 12:00	Next Generation Sequencing - Lee Murphy (E4.07)				
14:00 -15:30	Critical Evalution Skills 2 (E4.07)				
Monday 4th Nover	mber				
09:30 - 12:00	Biological Imaging - Ann Wheeler & team (E4.07)				
14:00 - 15:30	Degron tagging in human disease models: Opportunities and challenges - Andrew Wood (E4.07)				
Monday 11th Nove	ember				
09:30 - 12:00	Analysing Imaging Data - Ann wheeler (E4.07)				
14:00 - 15:30	Critical Evalution Skills 3 (E4.07)				
Thursday 14th No	vember				
15:00 - 17:00	POGS Thesis Writing Workshop				
Monday 20th Nove	ember				
09:30 - 12:00	Super resolution imaging - Ann Wheeler & team (E4.07)				
14:00 - 15:30	Genome Integrity disorders in the information age - Andrew Jackson (E4.07)				
Monday 25th Nove	ember				
09:00 - 12:00	Advanced Proteomics and Metabolomics - Alex von Kriegsheim, Jair Marques Junior (E4.07)				
14:00 - 15:30	Critical Evalution skills 4 (E4.07)				

Friday 30th November - Christmas Talks						
Monday 2nd Dece	Monday 2nd December					
09:30 - 12:00	Experimental Model Systems - Cameron Wyatt, Laura Lettice (E4.07)					
14:00 - 15:30	Computational Cancer Genomics - Colin Semple (E4.07)					
	ember - HGU Rotation Talks (E4.07)					
Monday 13th Janu	uary - 1st Rotation project write-up deadline - Start 2nd rotation					
Monday 13th Janu	Jary					
09:30 - 12:30	Translating your research - Helen Nickerson & Sarah Trewick (E4.07) TBC					
14:00 - 15:30	Critical Evalution Skills 5 (E4.07)					
Monday 20th Janu	lary					
10:30 - 12:00	Drug Development - Neil Carragher, Stefan Symeonides (E4.07)					
14:00 - 15:30	Regulation of genes and transcripts by the piRNA pathway - Ansgar Zoch (E4.07)					
Monday 27th Janu	Jary					
11:00 - 12:30	Biomedical Data Science - Catalina Vallejos (E4.07)					
14:00 - 15:00	Research Data Management Essentials - Steve Fox (E4.07)					
15:00 - 16:30	Critical Evalution Skills 6 (E4.07)					
Monday 3rd Febru	lary					
09:00 -12:00	Genome Engineering - Andrew Wood, Pleasantine Mill (E4.07)					
14:00 -16:30	Allele-aware functional genomics - Chris Ponting, Breeshey Roskams-Hieter - (MEC, Computing Lab)					
Monday 10th Febr	uary					
09:00 - 10:30	3D genome organisation. How to analyse it and determine its function? - Wendy Bickmore (E4.07)					
14:00 - 15:30	Critical Evalution Skills 7 (E4.07)					
Monday 17th Febr	uary					
09:00 - 10:30	Mechanisms of long-range gene regulation in development and disease - Hannah Long (E4.07)					
10:30 - 13:00	Alphafold Modelling (E4.07) TBC					
14:00 -15:30	RNA processing and gene regulation - Javier Caceres (E4.07)					
Monday 26th Febr	Monday 26th February					
09:00 - 10:30	Protein Variant Interpretation - Joe Marsh (E4.07)					
10:30 - 12:30	Scientific Graphics - Craig Nicol (E4.07)					
14:00 -15:30	Critical Evalution Skills 8 (E4.07)					
Monday 3rd March	Monday 3rd March					
09:30 - 10:30	The role of epigenetics in human disease - Duncan Sproul (E4.07)					
14:00 - 15:30	Innate immune signalling of self-nucleic acid in human disease - Yanick Crow (E4.07)					

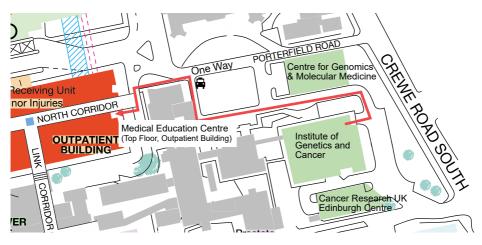
Monday 10th March					
09:00 - 12:00	Advanced Imaging Workshop - Ann Wheeler (E4.07				
14:00 - 15:30	Critical Evalution Skills 9 (E4.07)				
Monday 17th Marc	Monday 17th March				
14:00 - 15:30	Critical Evalution Skills 10 (E4.07)				
Monday 24th March					
09:00 - 10:30	Non-Coding mutations disrupt gene regulation and cause congenital defects - Laura Lettice - (E4.07)				
14:00 - 15:30	Chromatinopathies - Rebekah Tilotson (E4.07)				
Thursday 20th March - 2nd rotation project write-up deadline					
Friday 28th March - HGU Rotation 2 Talks					
Monday 31st March - HGU PhD Starts					
Monday 9th June - 10 week report write up deadline					
Thursday 12th June					
09:00 - 12:00	10 week report meeting (E4.07)				
Friday 13th June					
09:00 - 12:00	10 week report meeting (E4.07)				
Tuesday 26th August					
09:00 - 17:00	John Inglis Talks (Lecture Theatre)				

HGU Rotation Project Talks

Wednesday 11th S	September - IGC Lecture Theatre				
11:00 - 11:30	Joe Marsh - In silico mutational scanning to understand and predict protein function and genetic disease				
11:30 - 12:00	Wendy Bickmore, Luciana Gomwz-Acuna - Mechanisms of transcriptional activation by hormone-dependant enhancers				
12:00 - 12:30	Duncan Sproul - Using single molecule approaches to understand DNA methylation maintenance				
15:30 - 16:00	Rebekah Tillotson, Yanick Crowe - Using ATR-X syndrome-causing Mutations to Understand Protein Function				
16:00 - 16:30	Nick Gilbert - Understanding Transcription Noise in Cancer -await image and description				
16:30 - 17:00	Greg Kudla - High-Throughput Discovery of Disease Mutations by in vivo Deep Mutational Scanning				
Thursday 12th Se	ptember - E4.07				
09:30 - 10:00	Luke Boulter - Understating how changing cell states promote tissue regeneration and repair				
10:30 - 11:00	Liz Patton, Jenny Nichols - Developing Human Models for Melanocytes				
11:00 - 11:30	Ansgar Zoch - Understanding chromatin remodelling at piRNA-targete transposons				
11:30 - 12:00	Andrew Wood - How Do Human Disease Mutations Influence Targeted Protein Degradation?				
12:00 - 12:30	Nezha Benabdallah - Epistasis and Combinatorial Biomarkers in Cancer				
Thursday 19th Se	ptember - E4.07				
15:30 - 16:00	Tom Deegan - Mechanisms for Completing Genome Replication				
16:00 - 16:30	lan Adams - Manipulating Chromosomes to Prevent Errors in Mammalian Meiosis				
16:30 - 17:00	Colin Semple - The roles of tumour metabolic phenotypes in genomic instability				
Friday 20th September - E4.07					
15:30 - 16:00	Hannah Long - Investigating long-range enhancers and 3D genome topology at a human craniofacial disease locus				
16:00 - 16:30	Andrew Jackson - Stem cells in growth and ageing				
16:30 - 17:00	Catalina Vallejos - Biomedical data science -await image and description				



NHS Outpatients Building Computing Suite 1 Medical Education Centre, 3rd Floor



Institute of Genetics and Cancer Cancer Research UK Edinburgh Centre South Seminar Room S1.14



Institute of Genetics and Cancer



Assessment Guidelines for all students

Assessment Guidelines

PhD, MD, MScR assessment guidelines

During the course of your studies you will regularly be assessed. This will comprise writing reports, attending and presenting at thesis committee meetings and completing an annual review on EUCLID. For part time students assessments should happen every year and follow this format.

In the Institute our assessments are based on the CMVM guidelines and further information can be found on the CMVM wiki (http://edin.ac/2crLMTx)

Assessment reviews on EUCLID

All students need to complete an assessment review on EUCLID which will be signed off by you, your supervisors and postgraduate director. Over the course of your project you will complete an annual review to coincide with your 10-week, first year, second year and every subsequent year until you finish your studies. In some cases your thesis committee will decide that an interim meeting (e.g., half way through your second) or an additional meeting (e.g., at the end of the third year of a three year funded PhD) would be helpful. Please ensure your reports and feedback are uploaded onto EUCLID for sign-off. The online student portal (EUCLID) can also be used to record other important milestones in your training in Edinburgh and your supervisor may log individual meetings with you on this system.

Student reports

As a guide these are the reports required for different programmes.

	MSc by Research	3 year PhD	4 year PhD	MD
10 week report	\checkmark	✓	✓	✓
6 month report	✓			
1st year report		✓	✓	✓
2nd year report		✓	✓	
3rd year report			✓	

We will send out the hand-in dates of these assessments to all MSc/PhD students when they commence their studies.

10 week report assessment

This report should be concise (1000 words excluding title, references, abstract or figure legends). As this report is being written at the beginning of your studies, we are most interested in what you plan to investigate over the next year. The report should include:

- · Title, and the names of you and your supervisor.
- · An abstract of less than 100 words.
- Introduction that provides sufficient background information for the reader to understand the proposal and that puts the scientific question(s) into context.
- A section that states the scientific question(s) that are being asked and the aims of the project.

- · A short section on any progress made to date.
- A section describing your proposal for the next year's work.
- Figures can be added in any section to help describe the project or to show any data that you have obtained in the first few weeks of your project. Figure legends should provide succinct description of the figure.
- · Reference List.

On completion, the report should be uploaded onto EUCLID and submitted to the Graduate Research and Training team: student-admin@igc.ed.ac.uk. Following submission you will be given feedback in the form of an email and/or meeting (depends on programme). This is also a good time to plan the composition of your thesis committee (see below).

First Year Review:

- submitted at 9 month stage for PhD students
- submitted at 6 months for MScR students

The next assessment stage is the first-year review. This rigorous review is your opportunity to demonstrate your suitability to progress and will consist of three elements:

- · a written report from the student
- · a meeting with the student and thesis committee
- · a written report by the thesis committee

Student's written report: The report should adopt a logical format and be of a high standard. It should be typed and free of typographical and grammatical errors. A clear statement of the aims of the project should be included in addition to a brief account of methods and their validation. Whilst it is recognised that at this stage students may not have substantial data, preliminary results should be documented and interpreted with a clear statement of intent as to immediate future studies (these might be expected to form the basis of discussion at interview). The text should be referenced as for a scientific paper and references listed at the end of the report. It is expected that the report should be around 5000 words. It should be discussed with supervisors. It is often useful to ask your supervisor for an example report from a previous student. The student's report should be available to members of the thesis committee at least one week before the thesis committee meeting, allowing time for adequate consideration of the reports, and reports should be uploaded onto EUCLID.

Thesis Committee Meeting: This meeting will involve the student and thesis committee. The meeting is normally expected to include a short (10-15 minute) presentation by the student introducing the project, describing methodology and any preliminary results and identifying future studies. Students are strongly encouraged to rehearse with supervisors before the interview. You should expect the thesis committee to discuss specific points of content and organisation arising from the written report during the course of interview. You will have an opportunity to initiate a dialogue and, if necessary, raise matters of concern with the committee.

Feedback: The thesis committee should make an assessment of the student's written report, performance at interview and overall progress. The student should be informed of the committee's opinion during the meeting, they will then write a report, normally within one week of the meeting, summarising the assessment. Good and very good progress should be credited; any unsatisfactory aspects of performance should be clearly defined with an attempt to identify underlying reasons. It should make clear recommendations as to subsequent progress and action and be signed by all members of the committee. The student will have an opportunity to see the report, and be able to discuss strengths, weaknesses and any issues of concern with the chair in the absence of his/her supervisor(s). The student can also add comments before

signing the report. An unsatisfactory report may be used for future discussions or as the basis for re-registering students for a different degree or in very rare cases discontinuing studies (see outcomes). It is therefore essential that clear details of remedial action or the reasons for change in registration are documented. The signed thesis committee assessment should be uploaded onto EUCLID.

Outcomes: An initial recommendation will be made as to whether student progress is satisfactory or is inadequate in one or more aspects. In the case of inadequate performances a further recommendation from the thesis committee will be needed in terms of whether the student is (i) re-assessed or (ii) re-registered for a different degree, change in period of study or discontinued. In these cases it would be expected that students are totally unsatisfactory or severely deficient in several areas of their study.

Second Year Review

The second-year report does not need to be as long as the first year report but should contain a clear indication of achievable plans for the following year and an outline plan for the thesis. As for the first year review the student should organise a meeting with the thesis committee who will also write a report. Your second-year report and assessment from the thesis committee should be uploaded onto EUCLID.

Subsequent Reviews

For four year and continuing students there will be reviews every year until submission. Sometimes these will require a thesis committee meeting and this should be discussed with your supervisor.

Final Year Talk

Students in their final year will be scheduled to give a talk to their centre. These are a fun opportunity to present to your friends and colleagues and should be seen as an opportunity to showcase your work. These will be organised by student admin and your graduate director.

Thesis committee

The composition of the thesis committee will vary depending on your programme of study. It will comprise of your supervisors including a day to-day lab supervisor where appropriate, an external committee member and a Chair. The external may be from the same building, but should be independent of the supervisors. The Chair should be someone with experience of student supervision of at least Senior Lecturer level. For MScR and MD the roles of the chair and external are often combined.

General Information

- Postgraduate transferable skills programme
- Social media
- POGS
- Social committee
- Annual/sick leave
- Health and Wellbeing
- Pastoral Support Committees
- Annual Student / Supervisor
- Structured Discussion

Postgraduate transferable skills programme - Institute of Academic Development (IAD)

www.ed.ac.uk/institute-academicdevelopment

acquisition and development of The generic research and transferable skills is an important part of postgraduate training. Courses covering a wide range of skills are available to postgraduate research students in the Graduate School of Medicine & Veterinary Medicine through the transferable skills programme. This programme concentrates on the professional development of postgraduates, providing courses directly linked to postgraduate study (e.g. Thesis Workshop, Good Practice and Academic Paper Writing) and future careers (e.g. Successful Career Strategies for PhD Students, Local GRADschools). The programme also provides information on other training opportunities for postgraduates.

Courses are free of charge to postgraduate students in the College of Medicine and Veterinary Medicine. The programme has been designed to be as flexible as possible so that each student can tailor the content and timing of the programme to their own requirements. Most courses are run several times each year and last for between half a day and a day.

Workshops for postgraduate researchers by theme

The following workshops make up the core programme open to all postgraduate researchers, and are displayed by theme.

Research Planning and Management

- · Managing your Research Project
- Practical Project Management for Research Students
- · Viva Survivor
- · Innovation School
- · Managing your Research Data

Communication and Impact

- Designing Effective Slides
- Public Speaking, Networking and Engaging
- Poster Production
- Presenting made Easy Presentation Techniques
- Presenting Made Easy Delivering Presentations
- · Presenting your Poster Pitch
- Research, Researchers and the Media, a hands on approach to communicating your research

Writing and Publishing

- · Academic writing peer review
- · Beating Writers Block
- Developing a Writing and Publishing Strategy in the Internet Age
- · Effective Writing: Grammar
- · How to be your own best editor
- · Is my writing 'Academic' Enough?
- Just Write
- · Proof Reading
- Text, Coherence, Structure and Argumentation
- · The Writing Process: Getting Started
- · Writing a Literature Review
- · Writing Abstracts
- Writing Clinic
- · Writing for Publication
- · Writing Retreat
- · Writing Well: Language and Style
- · Academic Writer Creative Writer
- An Introduction to Copyright and Publishing
- This is what I do... and this is why it matters

Digital and Library Skills

- · Beginners Guide to Imaging
- Searching Literature and Managing Bibliographies
- · Managing a Bibliography in Endnote
- · Finding Academic Literature
- Social media for impact: strategy, connecting & metrics

Statistics

- Statistical Consultancy 1:1 Session
- · Introductory Statistics for Life Scientists

Personal Effectiveness

- · Conference and Events Organising
- · Creating Effective Collaboration
- · Creative Problem Solving for Researchers
- Imposter Syndrome: Why Successful people often feel like frauds
- Ease the Load Feel good about your busy life
- · How to be an Effective Researcher
- · Mapping your Mind
- Seven Secrets of a Highly Successful Research Student
- · Simply Assertive
- · Speed Reading
- Teambuilding and Leadership Fundamentals
- · Think Strategically Respond Rapidly
- Managing your Work, your Goals and Yourself

Public Engagement

- Communications Toolkit for a Public Audience
- Dialogue: Public Engagement Beyond Public Lectures!
- Storytelling Techniques for Effective Communication
- Voice and Presentation Skills Workshop
- How to Design a Public Engagement Process
- · Facilitation skills for public engagement
- · An Introduction to Public Engagement

Online learning

PhD student online training courses (topics include statistics; imaging; academic writing; and data management). Some you can do any time, and others run at specific times of the year.

- Statistics courses
- · Imaging for scientists
- Academic writing
- Research Ethics and Integrity an introduction
- · Data management training
- · Ready to research

The Edinburgh Local GRADschool is open to all PhD students in their final or penultimate year of study:

www.ed.ac.uk/institute-academicdevelopment/postgraduate/doctoral

Advice on using social media networks & confidentiality of information

Facebook, Twitter and other social media networks have changed the way we interact with each other and like them or not, they are a part of our society.

As some of you will carry out research where animals are involved, please ensure that you follow procedures to ensure our work continues to be ethical, credible and professional. Sharing images/discussions of animal work outside of the context of academic discourse is not appropriate. This not only applies to posts on social network sites but to informal discussions in the pub or on the bus.

Please remember you must not post the following information:

- Scientific research information, analysis, results or any other information and /or images relating to your work.
- Location details of research buildings where animal work is carried out.

Be mindful of your responsibilities

- Data Protection legislation do not disclose other people's personal information without prior permission.
- Be aware that any posts you make in a professional capacity (even private posts) are subject to data protection and freedom of information and may need to be disclosed.
- University policies apply: Students must not post materials about their work and locations if doing so would carry a risk to themselves and especially to others, including the University as an organisation (see section 5 University policies).

https://www.ed.ac.uk/students/healthwellbeing/wellbeing-services/disabilitysupport



POGS

The Postgraduate Society (POGS) is a student-run committee open to the Institute students from all years and centres. Our aim is to improve the student experience, promote collaboration, provide support and have fun! By organising events throughout the year we bring students together, helping them develop skills and career perspectives. Our most popular events include the annual student retreat, thesis writing workshop, pub quiz, poster evening, and careers event. All students are welcome to take part so don't hesitate to come say hi!

POGS is jointly funded by the Institute and the Deanery, which means (almost) all of our events are completely free! Joining the POGS committee is a great way to get involved with the Institute community, and have your say on how events are run. Meetings are held approximately once a month, and we are always looking for new committee members. To get involved, contact us at: pogs@igc.ed.ac.uk.





Best wishes, POGS



Vacation Leave

Students can take up to eight weeks' vacation time in a year, with agreement from their supervisor. There is no need to apply for an interruption of study when taking vacation leave.

Sick Leave

The policies on sick leave are evolving and depend on your funder. Please check information from your funding organisation or contact your programme director or Student Admin for advice.

Pastoral Support Committees

From September 2021 all students will be assigned a Pastoral Support Committee. This is completely independent of your thesis committee and will comprise two postdoctoral 'mentors' who will be based in different research teams and centres from you. The Pastoral Support Committee is there for to ask for advice, help, anything that you feel is not best addressed to your supervisor or thesis committee. The committee can meet as often as you like but at least once per year. Minutes won't be taken from the meetings but we will ask the committee to let us know when they have met.

Further information can be found on the IGC Graduate Research and Training website: <u>https://www.ed.ac.uk/institute-genetics-cancer/igc-graduate-research-and-training/information/</u>student-pastoral-support-committees

Student Support

The Institute of Genetics and Cancer is a family, looking out for each other. We are excited that you are becoming part of our family. If you need any local support a good place to start is with you supervisor. They will understand your situation and will want to look out for you. Alternatively please contact student admin (student-admin@igc.ed.ac.uk) or one of the postgraduate directors (Nick Gilbert, Val Brunton, Kathy Evans) and more information about different types of support is available at the back of this handbook.

Edinburgh university has lots of expertise in looking after students and a good place to start is the student Health and Wellbeing webpage: https://www.ed.ac.uk/students/health-wellbeing.

Code of Practice

https://www.ed.ac.uk/student-disability-service/staff/supporting-students/help-distressed-students

https://www.ed.ac.uk/files/atoms/files/copsupervisorsresearchstudents.pdf

Annual Student / Supervisor Structured Discussion

Many factors are important for a successful PhD. One of them is a good relationship between supervisor and student. To help this relationship it is important to be open and honest with each other and to manage each other's expectations. This is particularly important as the project progresses when there might be changes either due to the project or external factors. In our experience many problems can be avoided by having a dialogue, however this can be difficult for both parties, with neither wanting to put the other on the spot.

To overcome this problem, we would like all students and supervisors to have a "structured discussion" at the start of the PhD, and every year afterwards. In the first instance we will not monitor the results of the discussion, but instead ask students to let us know when you have had the meeting with your supervisor. In some cases, meetings will be online, in other cases face to face. Likewise for some PhD projects it might be relevant for all supervisors to participate, in other cases just the lead supervisor. Similarly, some topics will be relevant for different stages of your PhD.

https://www.ed.ac.uk/institute-genetics-cancer/igc-graduate-research-and-training/ information/annual-student-supervisor-structured-discussion

Register with a GP/doctor

It is important to make sure that you look after yourself whilst studying, both physically and mentally, and that you know how to get medical assistance if you need it.

The University has provided information about registering with your nearest doctor, known as a General Practitioner (GP).

Further information can be found on the New Students website here:

https://www.ed.ac.uk/students/new-students/ready-university/top-6-tasks/register-doctor



Student Health and Wellbeing

Feeling Good App



The Foundation for Positive Mental Health is Working with the University of Edinburgh to provide free access to the Feeling Good App.

www.ed.ac.uk/student-counselling/self-help/apps -podcasts-ted-talks-relaxation-recordings/feeling -good-app

Student Disability and Learning Support Service



Supports students with a range of health conditions, learning differences. disabilities and some temporary injuries.

www.ed.ac.uk/student-disability-service

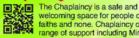
Advice Place



Professional, impartial and inclusive service for all students at the University of Edinburgh.

> www.eusa.ed.ac.uk/support and_advice/the_advice_place/

Chaplaincy



welcoming space for people of all faiths and none. Chaplaincy offers a range of support including Mindfulness, Yoga and the Listening Service.

www.ed.ac.uk/chaplaincy

SilverCloud



Online cognitive behavioural therapy. www.ed.ac.uk/student-coun selling/what-is-silvercloud

Membership to the Consent Collective



 Online support materials on consent. sex, gender, sexual harassment and relationships.

www.consentcollective.com/edinburgh

Student Counselling Service



Supports the mental health of all students at the University through short term counselling and referral to other support.

www.ed.ac.uk/student-counselling

University Health Centre

NHS General Practitioners who rent premises from the University and offer full G.P. services to patients.

EUSA Mental Health and Wellbeing Society



Provides an informal and friendly space where students can learn more about the importance of mental wellbeing. https://www.eusa.ed.ac.uk/

Togetherall



An online service offering self-help programmes, creative outlets and a peer support community monitored by mental health professionals.

www.togetherall.com/en-gb/

If you would like to discuss student health and wellbeing or any of the resources above, please contact: student-admin@igc.ed.ac.uk



MENTAL HEALTH DURING YOUR PHD

47%

NO MORE TICK BOXES

A study by the University of California, Berkeley, found nearly half of postgraduate students met criteria to classify them as depressed.¹

=

WHAT YOU MAY BE EXPERIENCING/FEELING (YOU ARE NOT ALONE, I PROMISE)

IMPOSTER SYNDROME

You got pretty good at doing essay and lab reports - they were all short term tasks. You also got good at figuring 7 in 1 e ne Someone is going to figure out you out what questions might be asked in exams. Now you I don't have what it takes to [do these experiments, write a thesis, succeed in academia]. These are all classic signs of nentality can be particularly tough. Tip: break down your imposter syndrome. Tip: reframe your thinking. Aim for progress not perfecti research into small, manageable goals. ISOLATION / GUILT FIRST TIME FAILING Writing your thesis can be a particularly lonely, isolating task. This can also be coupled with feelings You've always been the best student at school, and you did pretty well at university too. Now your science isn't of guilt when going about your daily life as "you working and everyone around you seems to be getting should be writing". Tips to manage this include stil attending research group meetings/departmental esigned to work. Tip: remember, you are at the forefront of scientific research - if it was easy it would already eminars whilst writing. This can also be coupled with writer's block'. Tip: when writing, start by making

have been done

COMPETITIVE LANDSCAPE

nately, academia often fo ld be the other way around. This is made worse nly way to gauge how well you are doing is to compare self against others. Tip: no two PhD projects are the same. so avoid comparing the

in these long hours. Tip: aim to be efficient inside normal working hours then focus on "you" time. A hard truth is only 7 in 200 PhD graduates become full professors.⁵ During your PhD, make sure to work on other "soft skills" as well as doing your research. Like making a poster for an online Twitter competition for e

55%

Δ

figures - it is far easier to write about what a figure means

THE WORK | LIFE STRUGGLE

There is an inherent culture of acceptance in

trait observed in academia, where peole work long

hours due to anxiety/stress, but are not being efficient

SELF-HARMING? ARE THOSE AROUND YOU STRUGGLING? HERE ARE SOME POSSIBLE WARNING SIGNS SUICIDAL THOUGHTS? CALL SAMARITANS NOW ί • ON 116-123 INCREASED DECREASED WORKING LOOKING INCREASED RFING .INKING OR EMAIL JO@SAMARITANS.ORG LONG HOURS DRINKING FATING FATING ABSENT DISHEVELLED SOME WAYS TO HELP MANAGE YOUR MENTAL HEALTH AND WELLBEING



A poster by Dr Zoe Ayres (not a medical professional). Free to distribute





Mental Health Portal

Do you want to learn about mental health and improve your resistance to stress, anxiety and other pressures?

Visit the IGC Mental Health Portal

- · Tips to improve Resilience
- Free resources for EVERYONE
- · Access at your own pace, in your own time, with no pressure or deadlines
- · Can be used to support others as well as yourself

IGC Mental Health Portal: https://edin.ac/mental-health-portal





Mental Health First Aiders

Are you concerned about your mental health or need someone to talk to?

The IGC Mental Health First Aid team are here to help!

The team are:

- Trained to provide a confidential listening service for ALL staff and students
- · Able to signpost to a range of different free resources, proven to help

For a full list of the IGC Mental Health First Aid Team visit: https://edin.ac/mental-health-first-aiders



Useful Links

Useful links

General

College PG Office contacts https://www.ed.ac.uk/medicine-vet-medicine/ postgraduate/contact-us/

College PG research wiki (includes PG handbook, software available to students etc.)

http://edin.ac/2crLMTx

Code of Practice https://www.ed.ac.uk/institute-academicdevelopment/postgraduate/doctoral/advicesupport/regulations

Assessment regulations

https://www.ed.ac.uk/files/atoms/files/2019postgraduateresearch.pdf

Transferable skills training and support www.ed.ac.uk/schools-departments/instituteacademic-development/postgraduate/ doctoral

Searching the literature/ bibliographic management

A tool for running daily searches http://pubcrawler.gen.tcd.ie/

A free online alternative to Endnote and Reference Manager www.zotero.org/

(note also that many journals have free apps for browsing abstracts).

Research Ethics

General www.pnas.org/content/86/23/9053.full.pdf

Image manipulation www.jci.org/articles/view/21717/pdf

www.cell.com/abstract/S0092-8674(06)00676-3

http://jcb.rupress.org/content/166/1/11.full

Writing papers, giving talks

Advice on writing papers www.nature.com/nature/journal/v467/n7317/ full/nj7317-873a

Useful advice ranging from lab techniques to giving talks and posters http://bitesizebio.com

The Advice Place, Potterrow Reception, EUSA 5/2 Bristo Square, Edinburgh EH8 9AJ Tel: 0131 650 2656 https://www.eusa.ed.ac.uk/

Advice Guides and Resources

Here you can read all of our advice guides. If you would like them in an alternative format, please contact us and we will do our utmost to accommodate this.

MRC Human Genetics Unit 4 Year Programme

- Introduction to programme
- · Projects available
- Rotation timeline

The first six months

The HGU PhD program is following an exciting and innovative format. You will spend the first 6 months on an intensive training period leading up to your final choice of PhD project. This period comprises taught courses, talks from individual group leaders about their work, teaching sessions on a variety of topics from technology to clinical research, journal club sessions which will give you a chance to hone your analytic and presentation skills, and 2 rotation projects. The detailed timetable can be found in the handbook.

The choice of rotation projects is up to you (available projects are listed at the end of this section) and you can approach any relevant group leader to discuss the projects. You will see that there is some time between rotations, giving you a chance to look around and choose a new lab. The only formal constraint is that vou must spend time in 2 different labs. You may find, of course, that another student has already been accepted and that the PI is only willing to take on one student (as is normally expected of PIs). If this happens then try again in the next rotation period: if there is a real clash then lan Adams will help but do please try and resolve things between yourselves in the first instance. Bear in mind that there is no formal requirement for you to choose a PhD project in a lab in which you have done a rotation project, the rotations are just a chance for you to try different labs and projects out.

Many of the group leaders welcome students coming to their lab meetings which is a good way of seeing life in labs other than the ones where you are doing rotation projects, but please be sure to make contact with the appropriate PI in advance.

The PhD

After 2 rotations you will choose a PhD project. We will have individual meetings with you to discuss your choices in the event of any clashes. No supervisor will be able to take on more than one student HGU students must choose projects within the HGU, but apart from this you can go to any lab within the available project section. It is up to you to discuss possible projects with PIs you are interested in: this is a dynamic process in which you should be fully engaged. Note that supervisors are not obliged to take you on, you need to ask whether they are willing, or whether they have other interested students and so on. If your research project involves the use of animals or human participants. work must not commence until the relevant Home Office project and personal licences have been awarded, and appropriate Local Ethical Approval Committee has been granted. We will not be producing PhD project outlines from supervisors. Rather, at the PhD 10 week stage (June) you will have to produce a short report that outlines the project that you will pursue. This will then be discussed and refined if necessary by your supervisors (more detailed guidelines are given under Assessment Procedures). You will then spend 3 years in the lab, winding up by April of your final year. You will then have a further 6 months to write up your thesis but remember it is imperative that you submit your thesis by the final university deadline of September of year 4!

We hope that this novel structure for PhD study will be as exciting for you as it has been for us to develop it. We will be asking for your feedback at several stages of the course - please feel free to air your views, and approach us about any issues you have, and help us to make the HGU PhD programme a huge success!

Nick Gilbert

Lab Rotations

Each student will do 2 rotation projects of around 3 months. Contact details and summaries of research interests of eligible supervisors are all given in this booklet (note there are some people unable to take students for rotations, please check), and during the first week you will be hearing research talks by some of these PIs.

The choice of rotation projects is up to you you are responsible for approaching potential supervisors to discuss their willingness to take you on and to jointly come up with a plan of work. Remember the project won't be formally assessed as part of your PhD, so make the most of your time to experience different techniques, and get a feel for life in different labs.

The only formal constraint is that you must spend time in 2 different labs. You may find, of course, that another student has already been accepted and that the PI is only willing to take on one student (as is normally expected of PIs). If this happens then try again in the next rotation period; if there is a real clash then one of us will intervene but do please try and resolve things between yourselves in the first instance. Bear in mind that there is no formal requirement for you to choose a PhD project in a lab in which you have done a rotation project, the rotations are just a chance for you to try labs out.

At the end of each rotation you have to write a report about your project, to be handed in by the end of the week after you finish in that lab. This should be in the format used by journals such as those on Biomedcentral, i.e. divided up into brief sections of background, results and conclusions and no longer than two sides of A4 (excluding figures).

This abstract should be submitted to the Institute of Genetics and Cancer PGSC by emailing:

student.admin@igc.ed.ac.uk

Supervisors will ask to meet up with you if there are any concerns.

Manipulating Chromosomes to Prevent Errors in Mammalian Meiosis

Supervisor: Dr Ian Adams

Rotation Oct-Dec Rotation Jan-March

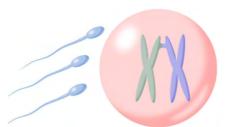
Inheriting the wrong number of chromosomes is one of the most common types of human genetic disease. These inherited aneuploidies arise particularly frequently in the oocytes of older mothers, and are typically caused by a failure to generate or maintain physical links between homologous chromosomes during meiosis. However, the molecular mechanisms involved in generating linked pairs of meiotic chromosomes then maintaining the resulting structures, usually for decades in human oocytes, remain poorly

Mechanisms of Transcriptional Activation by Hormone-Dependent Enhancers

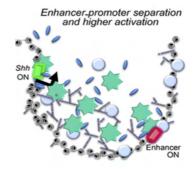
Supervisor: Prof Wendy Bickmore & Dr Luciana Gomez-Acuna

Rotation Oct-Dec Rotation Jan-March

How enhancers exert their action on their target genes over long genomic distances not fully understood. Our current is understanding of how enhancers function does not necessarily involve very close proximity of enhancer and promoter, but rather their engagement through transcriptional hubs We investigate the molecular mechanisms involved in enhancer-promoter communication with special interest in those leading to local chromatin decompaction, by deploying and developing cutting-edge techniques in advanced imaging, synthetic biology, molecular and cell biology. Using hormone-dependent enhancers in cancer cell line models, this project aims to uncover whether DNA damage, linked to increased transcription and the concomitant DNA-



understood in mammals. We will use geneedited mice and super-resolution imaging to build on our recent findings identifying new pathways involved in these processes with a view to manipulating the structure of meiotic chromosomes and chromosome-associated proteins to reduce chromosome segregation errors and prevent aneuploidies from arising in meiosis.



damage response, contribute to enhancerpromoter chromatin decompaction and proper transcriptional activation. This could point to an unexpected interplay between transcription and DNA-damage.

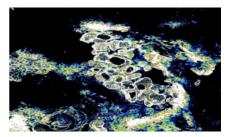
Benabdallah NS, Williamson I, Illingworth RS, Grimes GR, Therizols P, Bickmore WA. (2019). Decreased Enhancer-Promoter Proximity Accompanying Enhancer Activation. Molecular Cell 76: 473-484.e7.

Understating How Changing Cell States Promote Tissue Regeneration and Repair

Supervisor: Dr Luke Boulter

Rotation Oct-Dec Rotation Jan-March

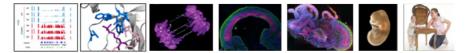
The adult liver is remarkably regenerative and yet lacks tissue stem cells. In order for epithelial structures in the liver to regrow following injury mature cells must lose some of their terminally differentiated characteristics and become more "progenitorlike". Using the adult bile duct as a model of tissue regeneration, we have identified a population of cells which become primitive during regeneration and sense their local environment to promote tissue regeneration. During this project we want to address two unresolved questions 1. Are these newly emerging cells essential for tissue regeneration and 2. How are they formed following injury.



Within rotation will а vou use immunofluorescent imaging to more closely characterise these cells and isolate them from tissue to determine whether they have more progenitor-traits than their differentiated counterparts. Further into the PhD project. you will work with transgenic animal lines, single nuclear RNA sequencing and high content imaging to dissect the role of these cells in regeneration and define whether this population of cells could be manipulated to promote regeneration in diseases where there is a bile duct deficit.

Stem Cells in Growth and Ageing

Supervisor: Dr Andrew Jackson Rotation Oct-Dec Rotation Jan-March



Our lab works on single gene human disorders with extreme phenotypes, including the smallest people in the world. As well as discovering new genes, we are interested in what these conditions can tell us about biological and common disease processes. To do so we use multiscale approaches including biochemistry, informatics, cell biology and animal models to discover new pathways and processes.

We have made recent unpublished discoveries which link epigenetics and stem cell function with growth and ageing. We are developing cellular models to understand these links, and the mechanism by which DNA methylation alters stem cell output, that may shed light on stem cell plasticity, and exhaustion, with translational relevance to regenerative medicine.

Developing Human Models for Melanocytes

Supervisor: Prof Liz Patton & Prof Jenny Nichols

Rotation Oct-Dec Rotation Jan-March

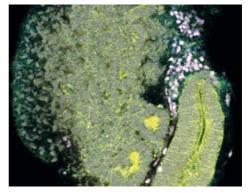
Melanocytes are pigment-producing cells that colour our hair, skin and eyes, and are the origins of melanocyte disease, including melanoma, mosaic melanocytic disease, vitiligo and albinism. Melanocytes also play less understood, but vital, roles in the ear, heart and other organs. We know little about how melanocytes develop in human embryos. Current models such as mouse and zebrafish provide critical insight into the developmental biology of these cells, however, they cannot fully explain human melanocyte disease. Using human gastruloids generated from stem cells derived from very early human embryos, this project will develop the first

How Do Human Disease Mutations Influence Targeted Protein Degradation?

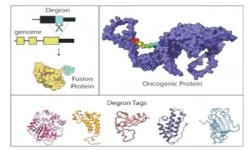
Supervisor: Dr Andrew Wood

Rotation Oct-Dec Rotation Jan-March

Despite recent advances our in understanding of human disease biology. many critical disease proteins remain beyond the reach of drug design. Targeted protein degradation provides a new approach to target such proteins using small molecules that accelerate the rate of destruction by the ubiquitin proteasome system. This project will use protein degradation biosensors in combination with genome editing, mutational scanning and protein engineering to better understand how common mutations in human oncogenes influence the activity of protein degrader drugs.



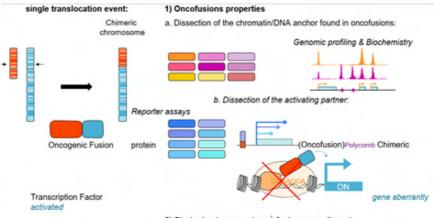
ever tools to study melanocytes in the developing human tissues in a dish, including fluorescent reporter lines and lineage tracing technologies. These studies will be combined with genomic studies to capture the molecular landscape and fate of melanocytes over time, and how these fascinating cells are altered by disease mutations.



Investigating the Role of Chimeric Fusion Proteins in Sarcoma

Supervisor: Dr Nezha Benabdallah

Rotation Oct-Dec Rotation Jan-March



2) Strategies to prevent oncofusions recruitment

Fusion-driven sarcomas are cancers of the bones and soft tissue that primarily affect young individuals. A key distinction of fusion-driven sarcomas, compared to other forms of cancer, is their genetic simplicity, typically driven by a single chromosomal translocation that fuses two previously separate transcription regulation proteins. The expression of an oncogenic fusion protein alone can induce cancer through the deregulation of target genes. In contrast, the removal of the fusion protein in sarcoma leads to reduced proliferation and cell death, underscoring their significance in cancer initiation and maintenance.

Our research aims to dissect the function of these oncofusions, which act as chimeric transcription factors, to advance our understanding of the underlying mechanisms of sarcomas, provide new avenues for therapeutic development, and enhance our knowledge of epigenetic cancer initiation. Moreover, many key fusion proteins contain poorly characterized chromatin regulators. By investigating these 'blind spots,' our research will contribute to a more comprehensive understanding of the human proteome.

By ectopically expressing oncofusions in human mesenchymal stem cells, we will investigate new and uncharacterized oncofusions within a consistent genetic background. Our study will focus on: 1) understanding the impact of chimeric transcription factors on gene regulation, and 2) designing biology-informed therapeutic strategies to target the mechanisms underlying these oncofusions.

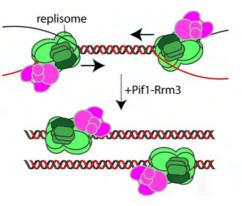
Mechanisms for Completing Genome Replication

Supervisor: Dr Tom Deegan

Rotation Oct-Dec Rotation Jan-March

DNA replication is driven by a molecular machine called the replisome, which is assembled at tens of thousands DNA replication origins in every cell cycle. Every time two replisomes emanating fro neighbouring origins converge and meet one another. DNA replication terminates leading to the local completion of DNA synthesis, and replisome disassembly. The faithful termination of DNA replication is fundamental for cellular fitness, as a failure to replicate even a short stretch of DNA would lead to chromosomal instability and/ or more mutagenic forms of DNA synthesis. This might be particularly relevant at Common Fragile Sites (CFSs), which are key hotspots for genome instability in human cancers that frequently fail to complete DNA replication before mitosis.

We have previously demonstrated that the Pif1-Rrm3 DNA helicases are specifically required to allow converging replisomes to pass one another during termination (Deegan et al. Mol. Cell, 2019). Our recent unpublished work has identified a new



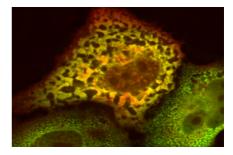
mechanism for how Rrm3 is recruited to replisomes. However, a molecular explanation for why accessory DNA helicases are required to prevent replisome stalling specifically during termination is lacking. This project will use in vitro DNA replication systems reconstituted with purified veast and human proteins, as well as cutting edge DNA sequencing and structural biology approaches, to understand the molecular basis for replisome stalling during replication termination. As well as exploring a fundamental and understudied area of chromosome biology, this project may also inform our understanding of numerous human diseases in which replication termination factors are mutated.

High-throughput discovery of disease mutations by in vivo deep mutational scanning

Supervisor: Dr Grzegorz Kudla

Rotation Oct-Dec Rotation Jan-March

Understanding which mutations lead to disease is a central goal of modern biology and medicine. Deep mutational scanning is a new approach that combines synthetic biology, next generation sequencing and computational analysis to systematically measure the effects of all possible mutations in a selected gene. So far, deep mutational scanning experiments were typically conducted in tissue culture. In collaboration with the Boulter and Khamseh labs, we will perform deep mutational scanning of tumour suppressor genes in an animal model of carcinogenesis, to explore the mechanisms of cancer formation in a physiological setting.



Investigating long-range enhancers and 3D genome topology at a human craniofacial disease locus

Supervisor: Dr Hannah Long

Rotation Oct-Dec Rotation Jan-March

Perturbation of gene regulation is central to many human genetic developmental disorders. In our lab, we study human facial development motivated by the wide diversity in facial appearance between individuals and the high frequency of birth malformations impacting the face. As our model, we focus on the SOX9 regulatory domain where noncoding mutations have been associated To explore with facial dysmorphology. mechanisms of disease, we leverage in vitro differentiation of facial progenitor (cranial neural crest) cells and have previously demonstrated that loss of extreme longrange enhancers (Long et al, 2020) and perturbation of 3D genome topology (Chen*, Long* et al. in preparation) can be implicated

in human disease. There are a number of exciting projects available in the lab. In one project, we will further explore the role of local 3D chromosomal structure in gene regulation. To do so, we will leverage human embryonic stem cells coupled with genetic engineering, reporter assays, genomics, and chromosomal imaging to explore the role of novel structural features in gene regulation across craniofacial development at loci

implicated in craniofacial disease.

How does 3D genome topology contribute

to extreme long-range enhancer function?

cohesin • CTCF enhancer cluster

Exploring gene regulation in human

facial development and disease

Understanding Transcription Noise in Cancer

Supervisor: Prof Nick Gilbert

Rotation Oct-Dec Rotation Jan-March

Pathways have been identified, for many types of cancers (especially solid ones) curation remains rare. In many cases, this is due to the development of resistance to therapeutics, which arises due to the tumor cells' extraordinary capacity for genomic adaptation, which is commonly referred as plasticity [1]. Plasticity is one of the most crucial hallmarks defining what cancer is: indeed, throughout cancer progression, tumor cells have to continuously adapt to unfavorable conditions, be it hypoxic microenvironment, immune system attack, or chemotherapy[2]. A number of recent discoveries suggest that transcriptional plasticity provides the mechanism to this adaptation, and current thinking suggests that this is intimately linked to 3D structure [1,2].

In this project we will study transcription at the single cell level for both healthy and diseased cancer cells, with a view of quantifying transcriptional plasticity. We will use an interdisciplinary approach where we will draw from computational biophysics studies of chromatin folding [3] which can be linked to transcriptional output at the single cell level [4], providing a mechanistic underpinning for our expeirments. We will set up a cycle where experiments are compared with predicted mechanistic modelling to refine this, and modelling in turn generates hypothesis which drive the design of new experiments to test them.

Specifically, in this project we aim to obtain and analyse scRNA-seq and RNA-FISH data for transcriptional activity in single cells, in healthy tissues and in a variety of cancer tissues. We will analyse the transcriptional activity and heterogeneity in these samples, using TT-seq and computer simulations (which are predictive and can output transcription readout for each regulatory element in the human genome[3,4]) as cross-validation. Plasticity will be quantified with a series of observables, inspired by



information theory and statistical physics. For instance we will explore the link of entropylike quantities (as a measure of structural and transcriptional diversity, and of transcriptional noise) and plasticity assessed as susceptibility to environmental perturbation, such as subjecting cells to chemotherapeutic agents, which can in principle be studied both experimentally (by scRNA-seq of cells after treatment), and computationally (by using the HiP-HoP model of [3,4] based on epigenetic tracks on healthy and diseased cells, with and without treatment). Our combined measures provide a multifaceted assessment of the so-far quantitatively elusive concept of plasticity, which we believe will be of high interest outside the topic of transcriptional heterogeneity in cancer, providing the student working on the project with transferrable skills in a key area of research.

The student will gain training on singlecell molecular biology and sequencing experiments, such as scRNA-seq, RNA-FISH as well as other single-cell assays (such as ATAC-seg and other epigenetic mark analysis). Additionally, they will be exposed to bioinformatic and computational pipelines to analyse data with state-of-the-art codes aimed at studying transcription in single cell to extract noise and gene-gene correlations besides averages. An important training outcome will be the ability to interact with modellers and computational biophysicists. which is necessary in contemporary teams wishing to explore quantitatively biomedical problems.

[1] Househam, J., et al. Phenotypic plasticity and genetic control in colorectal cancer evolution. Nature (2022).

[2] Virk, R.K.A. et al. , Disordered chromatin packing regulates phenotypic plasticity. Science Advances 6, eaax6232 (2020).

In Silico Mutational Scanning to Understand and Predict Protein Function and Genetic Disease

Supervisor: Prof Joe Marsh

Rotation Oct-Dec Rotation Jan-March

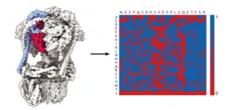
Recent advances in machine learning and the development of high-throughput deep mutational scanning strategies are revolutionising our ability to interpret human genetic variation. This project will use state-of-the-art variant effect predictors and protein structural models to perform in silico saturation mutagenesis on a variety of proteins implicated in human genetic disease. This will enable us to better understand protein function and its relationship to disease, and establish optimal strategies for computationally predicting the effects and molecular disease mechanisms of novel variants, thus ultimately improving the diagnosis and treatment of human genetic disorders.

Statistical and ML approaches to cluster longitudinal trajectories in molecular and clinical data

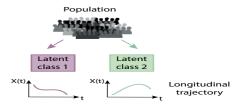
Supervisor: Dr Catalina Vallejos

Rotation Oct-Dec Rotation Jan-March

Longitudinal data plays an important role in biomedical research. Examples cover a wide range of applications: from the monitoring an individual's health status based on their measured biomarker values, to quantifying gene expression patterns along cellular differentiation trajectories. One important problem in this context is to identify aroups (e.g. of individuals, of genes) that share similar longitudinal trajectories. Several longitudinal data clustering techniques have been developed for this purpose. but there are limited benchmark studies. comparing their performance. This project aims to evaluate the performance of a variety of statistical and ML approaches for longitudinal data clustering using synthetic and real datasets. The goal is to develop



Gerasimavicius L, Livesey BJ & Marsh JA (2022) Loss-of-function, gain-of-function and dominant-negative mutations have profoundly different effects on protein structure. Nature Communications 13:3895



a reproducible benchmark pipeline that can be inform potential users and that can support the development and evaluation of new methods. By the end of the project, the student will be familiar with a wide range of longitudinal data clustering techniques as well as with best practice reproducibility workflows.

This project is entirely computational and requires R programming skills.

Using single molecule approaches to understand DNA methylation maintenance

Supervisor: Dr Duncan Sproul

Rotation Oct-Dec Rotation Jan-March

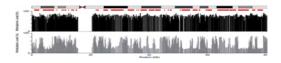
The failure to maintain developmentally established patterns of the epigenetic mark DNA methylation is a hallmark of cancer and aging. Our current understanding of the impact this failure is hindered by our lack of knowledge regarding the fundamental molecular mechanisms underpinning DNA methylation maintenance. We have developed single-molecule methods to measure DNA methylation patterns and techniques to introduce synthetic epialleles into cells. Together, these allow us to quantitatively track the evolution of DNA methylation patterns over time. This PhD

Understanding Chromatin Remodelling at piRNA-targeted Transposons

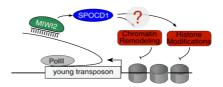
Supervisor: Dr Ansgar Zoch

Rotation Oct-Dec Rotation Jan-March

We study how the mammalian germline defends genome integrity against jumping genes called transposons. The germline, the cell lineage that will eventually make sperm and egg cells, passes on our genetic information to the next generation. Maintaining the integrity of the germline genome is thus paramount to the continuation of life. In mammals, germ cells undergo a reprogramming event that erases most epigenetic information and reactivates transposons, creating existential an challenge. Transposons threaten genome integrity through mutagenic transposition and deregulation of chromatin. The piRNA pathway identifies active transposon loci and targets them for epigenetic silencing by de novo DNA methylation. Yet, the mechanisms of how target-recognition induces silencing are not understood. We recently discovered



project seeks to use these approaches to dissect why DNA methylation patterns are not maintained in cancer and aging and can be approached from a laboratory or computational perspective.



essential effector proteins required for piRNA-mediated DNA methylation. We also demonstrated the relevance of transposon repression by the piRNA-pathway for human fertility. In this project we will build upon these discoveries to investigate how the piRNA pathway interacts with chromatin remodeller complexes to regulate transposon activity using a range of molecular biology tools in cell culture models.relevant to a broad range of diseases.

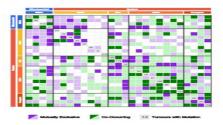
The Roles of Tumour Metabolic Phenotypes in Genomic Instability

Supervisor: Prof. Colin Semple & Dr Stuart Aitken

Rotation Oct-Dec Rotation Jan-March

High grade serous ovarian cancer (HGSOC) is an archetypal tumour type showing extreme genomic instability. We have discovered that HGSOC patients' tumours also often acquire mitochondrial (mt) mutations predicted to alter mt metabolism, based upon a large (N=324) tumour cohort with deep whole genome sequencing and RNAseg. These patients have significantly poorer survival but the mechanisms linking mt mutations to clinical outcomes are unknown (Ewing et al. 2024). It has long been known that disrupted mt function can cause the accumulation of the metabolite lacate, aiding tumour proliferation (Li et al, 2022), as well as efficient DNA repair (Chen et al. 2024). Thus it is possible that in HGSOC mt dysfunction may lead to metabolic phenotypes that underpin tumour aggressiveness and prime tumours for resistance to genotoxic agents, such as cisplatin and radiotherapy, resulting in poorer survival

Analysis of pan-cancer metabolomic and expression data has identified genemetabolite interactions (GMIs) linking gene expression patterns to the levels of particular metabolites in tumour samples (Benedetti et al, 2023). Recently it has been shown that GMIs can be exploited to impute otherwise unmeasured metabolite levels in tumours using a novel Bayesian appraoch to process bulk RNAseg data (Xie et al, 2024). In this project we will use the same approach to infer the metabolic phenotypes present in our HGSOC cohort and determine whether they are associated with differences in patient survival and genomic insability, as reflected in tumour mutational burdens. We will address the following aims.



1. Identify the range of metabolic phenotypes in HGSOC tumours based upon imputation from RNAseq data, and identify those associated with the presence of mt mutations.

2. Determine the association of metabolic phenotypes with tumour mutational burdens, complex structural variation and DNA repair phenotypes.

3. Determine the association of these metabolic phenotypes with patient outcomes.

The project would best suit a bioinformatics student with an interest in cancer genomics or an enthusiastic biologist keen to develop bioinformatics skills.

References

Ewing et al. 2024. Divergent trajectories to structural diversity impact patient survival in high grade serous ovarian cancer. bioRxiv doi: https://doi.org/10.1101/2024.01.12.575376

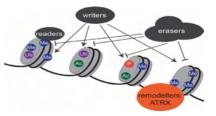
Using ATR-X syndrome-causing Mutations to Understand Protein Function

Supervisor: Dr Rebekah Tillotson and Professor Yanick Crow

Rotation Oct-Dec Rotation Jan-March

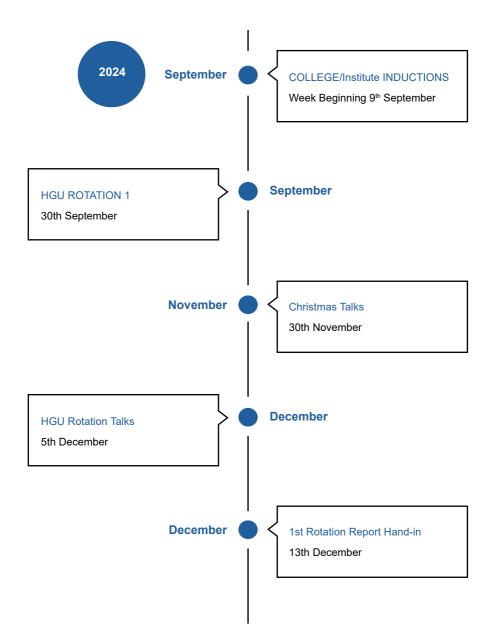
Chromatin factors write/read/erase epigenetic marks or remodel chromatin structure to regulate chromatin state and control transcription. These factors are essential for normal development, with mutations that disrupt their activity or alter dosage resulting in the so-called "chromatinopathies" – monogenic disorders affecting multiple tissues, almost invariably including the brain.

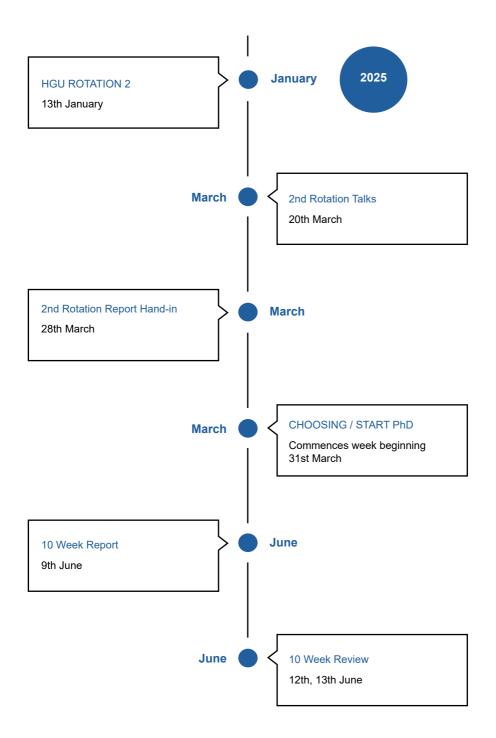
ATR-X syndrome affects males due to hypomorphic mutations in the X-linked ATRX gene, encoding a chromatin remodeller that has been implicated in gene regulation. How different classes of disease-causing mutations affect the domains within a protein can help us understand its molecular function. Notably, in ATR-X syndrome, missense/ in-frame mutations cluster in the chromatin binding and ATPase domains of ATRX.



There are several potential projects available in the laboratory, including one to interrogate a highly novel link between ATRX dysfunction and innate immune system induction. Projects will make use of cellular and mouse models and involve experimental techniques such as CRISPRediting, transcriptomics (RNA-sequencing and qPCR), analysis of chromatin binding patterns, co-immunoprecipitation, western blotting and fluorescence microscopy and well as bioinformatic analysis.

Training Timeline 2024 - 2025





INSTITUTE OF GENETICS & CANCER

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A new Facebook Group has been created for current on-programme students at the institute. This online space is a closed group and has been created specifically for students (not staff) for announcements, course materials, discussions and a place to get to know each other.

Join by searching Facebook for OFFICIAL IGC Students or visiting: www.facebook.com/groups/ OFFICIALIGCStudents

Do I have to join the group?

Yes. We hope the group will make life easier for everyone by having all the right information and people in the same place, reducing email traffic and providing a place for resources, questions and answers.

What if I'm not on Facebook or don't want to use my personal profile to join?

That's ok – contact us and we can help you set up a new profile, just for life at the Institute.

What is a closed group?

Only approved members of the group can see who the current members are and view posts in the group. Anyone on Facebook can see the group's name and description, find it through search and request to join (requests are approved or declined by Administrators), but they can't see any of the content or members.

Who will be in the group and who moderates it?

All postgraduate students on programme at the Institute.

Pauline and Alana are the Group Administrators with the Communications Manager as Moderator. Look out for group announcements from the Administrators – these flag key information. Join requests are approved by the Administrators, so no 'outsiders' will be able to join the group.

Can we say what we want?

Although this is your group, remember that the group represents the Institute and we expect members to behave as professionally as they would in person on campus. Inappropriate posts will be moderated and removed.